

Growth Performance of African Catfish (*Clarias Gariepinus*) Fed on Diets Containing Black Soldier Fly (*Hermetia Illucens*) Larvae Under Aquaponic System

Benard Maranga^{1,*} , Robert Kagali¹ , Kevin Mbogo² , Paul Orina³ , Jonathan Munguti⁴ , Eric Ogello⁵ 

¹Jomo Kenyatta University of Agriculture and Technology, Zoology, Nairobi, Kenya

²Jomo Kenyatta University of Agriculture and Technology, Biochemistry, Nairobi, Kenya

³Kenya Marine and Fisheries Research Institute, Aquaculture, Kisii, Kenya

⁴Kenya Marine and Fisheries Research Institute, Aquaculture, Sagana, Kenya

⁵Maseno University, Fisheries and Natural Resources, Maseno, Kenya

How to cite

Maranga, B., Kagali, R., Omolo, K., Orina, P., Munguti, J., Ogello, E. (2023). Growth Performance of African Catfish (*Clarias Gariepinus*) Fed On Diets Containing Black Soldier Fly (*Hermetia Illucens*) Larvae Under Aquaponic System *Aquaculture Studies*, 23(5), AQUAST910. <http://doi.org/10.4194/AQUAST910>

Article History

Received 03 March 2022

Accepted 13 November 2022

First Online 16 November 2022

Corresponding Author

Tel.: +254727669158

E-mail: marangabenard@gmail.com

Keywords

Black soldier fly

Fishmeal

Catfish

Growth performance

Abstract

Cost of fish production can be reduced by replacement of high-priced fishmeal (FM) with insects sourced ingredients. Four months feed experiment was conducted at a fish farm in Baringo County, Kenya to investigate effects of substituting fishmeal (FM) with black soldier fly larvae meal (BSFLM) on survival and growth performance of *C. gariepinus* under aquaponic system. Three test diets 35% crude protein content (CP) in which FM was substituted by BSFLM at 25%, 50% and 75% were formulated and experimented with commercial diet of 35% CP. Four weeks old *C. gariepinus* were stocked in 12 tanks at a density of 50 fish/tank and subjected to the diets. Fish were sampled every three weeks; water parameters were sampled weekly and mortality recorded on occurrence. Diet with 50% BSFLM obtained better FCR for formulated diets with no significance ($P < 0.05$) for FCR and survival. Weight gain of control diet (97.07 g) was significant ($P < 0.05$) compared to formulated diets 64.09g, 69.78g and 67.77g for 75%, 50% and 25% of BSFL replacement respectively. Growth performance and survival demonstrated that BSFLM has potential to substitute FM up to 75%. The fish productivity can be improved and feed cost reduced by incorporating fully defatted BSFLM with CP higher than 25.3% used for the diets.

Introduction

Globally, aquaculture production is the most immediate solution to declining capture fisheries, however, it also faces a number of challenges that prevent it from reaching its full potential (Barbu *et al.*, 2016). Outdated culture systems and high cost of fish feeds resulting from competition of raw materials have been identified as major setbacks in the growth of aquaculture in most developing countries. Modern aquaculture technologies such as aquaponic system offer an opportunity for exploiting limited land and scarce water to farm fish and crops within the same

system (Rurangwa and Verdegem, 2014) thus accelerating productivity. However, this requires innovative approach that use alternative sources of energy such as solar power, thus a shift from the high-cost national gridline electricity (Badiola *et al.*, 2018) characterized by power outage and high costs per unit of use. Aquaponic system offers an excellent alternative to traditional aquaculture practices that have struggled to intensify productivity. It is characterized by intensive fish production unit using a series of water treatment steps that include plants and filters for removing nutrients and impurities from water to facilitate its reuse (Espinal and Mutalic, 2019). Nile tilapia

(*Oreochromis niloticus*) and African catfish (*Clarias gariepinus*), are the most cultured species in Africa and have in previous studies demonstrated their candidature in aquaponics system (Ekawati *et al.*, 2021; Love *et al.*, 2015). A variety of plants can be grown within the aquaponic system under the hydroponic unit for nutrients uptake thus maintaining the integrity of water within the system for fish culture. Some of the plants grown under aquaponic systems include; Leafy vegetables (Spinach and Amaranth), Fruit vegetables (Cucumber and Eggplant), Root vegetables (Beets and Radishes) among other plants (Kim 2018).

C. gariepinus preference in aquaculture is as a result of its ability to tolerate high environmental temperatures and low dissolved oxygen thus making it a good candidate for intensive farming, (Safran, 2009). Under best management practices, *C. gariepinus* can be cultured intensively (Somerville *et al.*, 2014) under limited land and water resources (Huntington *et al.*, 2017). Demand for *C. gariepinus*, both for food and as bait in capture fisheries has been increasing substantially in Kenya in the last few years (Omondi *et al.*, 2001). Kenya's State Department of Fisheries and the Blue Economy estimates a demand of about 10 million *C. gariepinus* fingerlings per year for aquaculture and 18 million fingerlings per year for Lake Victoria capture fisheries. This adds up to a total demand of about 28 million *C. gariepinus* fingerlings annually, thus informing its selection for this study.

Quality, availability and cost of fish feeds are the most prohibitive aspects in Kenya's aquaculture development (Munguti *et al.*, 2021). Fish feeds are important component in the value chain of *C. gariepinus* production. Substantial percentage of *C. gariepinus* production cost is incurred in procurement of feeds (Adewolu *et al.*, 2010). Commercial feeds are generally expensive and their prices continue to rise due to competition for ingredients by other animal feed manufacturers, (FAO 2014). Complete diets contain plant and animal sourced ingredients that provide all nutritional requirements of fish should be formulated so as to achieve good health and maximum growth of cultured fish (Al Mahmud *et al.*, 2012). To realize economic viability, supply of highly nutritious and low-cost ingredients should be used for formulation (Munguti *et al.*, 2009). Therefore, it is important to replace the commonly used ingredients that are expensive with non-conventional locally available inexpensive ingredients to reduce the cost of fish feeds (Chepkirui *et al.*, 2011; Liti *et al.*, 2002).

Fishmeal is a conventional ingredient used for animal feeds formulation (Walker, 2009), with aquaculture industry accounting for 68% of fishmeal consumption in the world (Naylor *et al.*, 2010; Miles and Jacob, 2011). As such, market supply of fish meal is declining and becoming unsustainable due to over exploitation from nature making it very expensive for small scale fish farming (Munguti *et al.*, 2014). Therefore, diversification of protein sourced ingredients

for formulating fish feeds will result to significant reduction of the cost of fish production thus encouraging more farmers to take up fish farming (Ngugi *et al.*, 2007). The potential of insect-based ingredients in replacing fishmeal has shown positive results in growth performance on several fish species (Stamer *et al.*, 2014). Thus, insect meals have been incorporated in fish feeds formulation as protein source ingredient. Among the promising insect species for commercial feed formulation include the black soldier fly (BSF), common house fly, silk worms and house cricket (Tilami *et al.*, 2020). Termites and grasshoppers are also viable in animal feeds formulation though to a lesser extent. Recent studies have shown black soldier fly larvae to be a potential protein source and alternative to fish meal (FM) in formulating fish diets (Matteo *et al.*, 2020). The BSF larvae can easily be cultured using a variety of organic waste generated within the farm making it an eco-friendly insect and very cheap source of protein, hence a prospective ingredient to reduce production cost (Tran, Gnaedinger and Merlin 2015). Proximate composition of the larvae has shown a higher crude proteins content value of up to 45%, presence of microelements, amino and fatty acids in required quantities essential for fish growth (Xiao *et al.*, 2018). The present study was aimed at formulating three experimental diets that were tested against a commercial diet with percentage proportion of FM and BSFLM in formulated experimental diets gradually being reduced and increased respectively. The four diets were evaluated on survival and growth performance for intensive production of *C. gariepinus* reared under a solar powered aquaponic system.

Materials and Methods

The study was carried-out over four months duration at a private fish farm in Baringo county, Kenya (0°39'N, 36°05'E), within an altitude of 970 meters above sea level, average annual temperature and rainfall is estimated at 33°C and 684mm respectively. Proximate analysis of feed ingredients and formulated diets was conducted at food science laboratory of Jomo Kenyatta University of Agriculture and Technology (JKUAT) while chemical water parameters were analyzed at Kenya Marine and Fisheries Research Institute KMFRI laboratory, Baringo station.

Experimental Design and Set-up

The study was carried out under complete randomized block design using aquaponic system (Figure 1). The system comprised; 12 fish rearing tanks for fish culture, 4 sumps that had submersible water pumps for water recirculation and 4 hydroponic units that were used in growing of spinach (*Spinacea oleracea*) for nutrients uptake from fish rearing water before being recycled to fish rearing tanks.

Sourcing Ingredients, Proximate Analysis and Diet Formulation

Wheat bran (WB), cotton seed cake (CSC) and sunflower seed cake (SSC) were sourced from animal feeds raw material outlet within Nakuru town, partially defatted 2 weeks old BSFL cultured using kitchen waste was sourced from International Centre of Insect Physiology and Ecology (ICIPE) Nairobi while fish meal was obtained from Kendu Bay in Homa Bay County. Proximate composition of individual ingredients and diets were analyzed for dry matter (DM), crude protein (CP), crude fibre (CF) ether extract (EE) and ash content. The analysis was carried out in triplicate using the procedure described in AOAC (2005) and presented as shown in Tables 1 and 3 respectively, while percentage ratio of each ingredient in diets is presented in Table 2. (NFE) was determined by the difference method (DM – CP – EE –CF –Ash). NB: Commercial diet (D_c) 35% CP (control) was sourced from (Unga Farm Care Limited) within Nakuru town.

The ingredients were sun dried, ground and three test diets (D₁, D₂ and D₃) of 35% crude proteins (CP) content were formulated using Pearson Square method by substituting FM with BSFLM in (D₁, D₂ and D₃) at (50-50, 75-25, and 25-75) % ratios respectively. Using water, the formulated diets were made into dough and pelletized through 2 mm dice, then sundried appropriately before being packaged and stored at room temperature in plastic containers.

Stocking and Feeding

Clarias gariepinus fingerlings were obtained from Kenya Marine and Fisheries Research Institute, Sagana Aquaculture Research Center located in Central Kenya. Since aquaponics entails an intensive culture system, small area is used to culture a higher number of fish. A total of 50 *C. gariepinus* fingerlings weighing (5 ± 0.5g) were acclimatized for a week and thereafter stocked in 12 (1000 litre) tanks at a stocking density of 0.05 fish/litre of water. The three test diets and control diet (D₁, D₂, D₃ and D_c) were randomly allocated to the first four set of tanks and replicated thrice. Cover nets were placed over the tanks to control predators. Fish were hand fed twice daily at 09:30hrs and 15:30hrs at 5% and 3% body weight for the first two and the last two months of the trials respectively.

Fish Sampling and Growth Determination

20 litres buckets were halfway filled with 10 litres of water in which fish were placed after being removed from fish rearing tanks using a scope net. Length was determined using a metre rule bound on a board while weight was determined using weighing machine, WTC 2000. Sampling of fish was done after every three weeks to monitor growth of fish that was used to adjust the feeding rate.

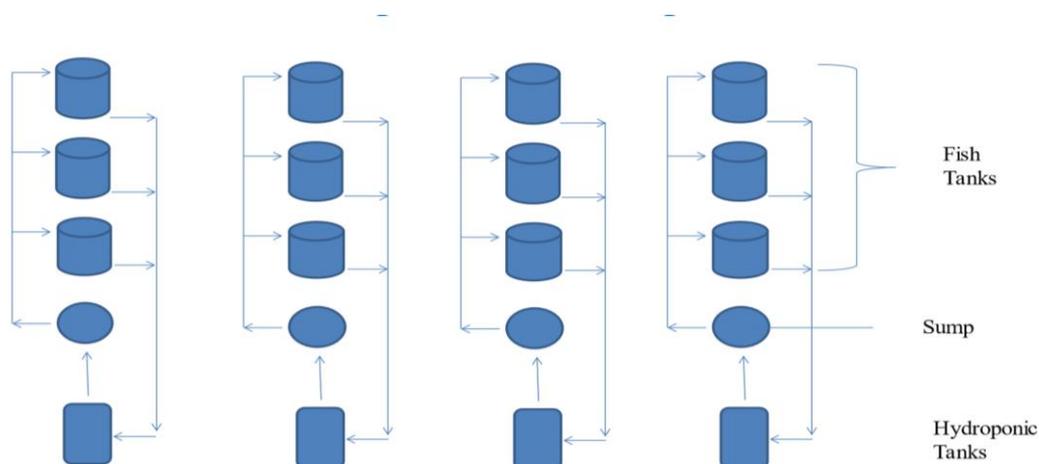


Figure 1. The experimental aquaponic system (Not drawn to scale). The arrows indicate the direction of flow of water within the system.

Table 1. Percentage (%) proximate composition of analyzed ingredients used in diet formulation

Parameter	DM	CP	EE	CF	Ash	NFE
BSFL	95.8	25.3	27.3	8.0	14.7	20.5
FM	89.94	48.80	12.4	1.28	9.28	18.18
CSC	92.14	32.29	8.60	6.46	18.54	26.25
SSC	91.85	26.83	9.90	16.92	9.44	28.76
WB	90.08	17.10	5.01	10.96	8.43	48.58

DM: Dry matter, CP: Crude proteins, EE: Ether extract, Crude fibre: NFE: Nitrogen free extract.

BSF: Black soldier fly larvae, FM: Fishmeal, CSC: Cotton seed cake, SSC: Sunflower seed cake and WB: Wheat bran

Water Quality Analysis

Specified water quality parameters (dissolved oxygen, temperature, pH, conductivity and total dissolved substance) were monitored after every week at 07:00hrs using a multiparameter water quality meter (YSI) while water samples were collected in 200ml sample collection bottles and taken to the laboratory to determine the levels of ammonia, phosphorous, nitrites and nitrates using UV Shimadzu spectrometer (UV-1800).

Evaluation of Survival and Dietary Performance

At the end of the experiment, fish were harvested, counted, weight and length documented. Survival, Growth and feed efficiency were evaluated by the following standard formula.

$$\text{Survival rate (SR) (\%)} = 100 \times \frac{\text{Final number of fish}}{\text{Initial number of fish}}$$

$$\text{Daily growth (DG) (g)} = \frac{\text{Final weight}}{\text{Time (Exp days)}}$$

$$\text{Body weight gain (BWG) (g)} = \text{Final weight} - \text{Initial weight}$$

$$\text{Specific Growth Rate (SGR) (\%)} = 100\% \times \left[\frac{\ln \text{Final weight (g)} - \ln \text{Initial weight (g)}}{\text{Time (Exp days)}} \right]$$

$$\text{Fish Food Conversion Ratio (FCR)} = \frac{\text{Feed provided (g)}}{\text{Weight gain (g)}}$$

Data Analysis

Data storage and management was done using Microsoft Excel spreadsheet for windows 2010 while analysis was done using SPSS Version 23.0 for windows. Means for growth and water quality parameters for all the treatments were compared using Analysis of Variance (ANOVA). Significant difference for all inference tests were separated using the Tukey-HSD post hoc, at 0.05 level of significance. The data was presented using tables and a graph plotted using excel spreadsheet.

Results

Fish Survival, Feed Utilization and Growth Parameters

Survival of fish under the set growth conditions and experimental diets was above 95% with relatively low mortality rate experienced in the first week of stocking especially for fish cultured under diet 3 (D3) (Table 4). There was no significant difference (P<0.05) observed in survival rate (SR) among all the dietary treatment and control diet under the present study. Fish that were fed on diets that BSFLM substituted FM had statistically

Table 2. Percentage (%) crude proteins content, composition of ingredients and essential amino acids in test diets containing varying inclusion of BSFL in partial substitution of FM

Parameter	Unit	Experimental diets			
		D ₁	D ₂	D ₃	D _c
Proteins analysis					
Crude proteins	(g 100g ⁻¹)	35.0	35.0	35.0	35.0
Ingredients					
BSFL	(g 100g-1)	13.75	6.875	20.625	-
FM	(g 100g-1)	13.75	20.625	6.875	5.7
Soybean meal	(g 100g-1)	-	-	-	25.05
Cotton seed cake	(g 100g-1)	16.27	16.27	16.27	-
Sunflower seed cake	(g 100g-1)	25.495	25.495	25.495	-
Wheat bran	(g 100g-1)	30.735	30.735	30.735	68.15
Essential amino acids					
Methionine	(g 100g-1)	-	-	-	0.5
Lysine	(g 100g-1)	-	-	-	0.2
Choline	(g 100g-1)	-	-	-	0.4

D₁=Diet 1, D₂=Diet 2, D₃=Diet 3 and D_c=Control diet

Table 3. Proximate composition of formulated test diets containing different percentages of BSFL as a substitution of FM

Parameter	Unit	Means ± SD			
		D ₁	D ₂	D ₃	D _c
Dry matter	(g/100g)	91.43±0.81a	91.57±0.35a	91.57±0.09a	91.55±0.63a
Crude proteins	(g/100g)	35.39±0.07a	35.28±0.10a	35.30±0.02a	35.41±0.19a
Ether extract	(g/100g)	9.09±0.14b	10.28±0.10c	11.47±0.93d	7.25±0.32a
Ash	(g/100g)	10.65±0.05b	10.05±0.06b	6.65±0.08a	10.31±0.06b
Crude fiber	(g/100g)	5.36±0.39a	6.20±0.52b	6.54±0.18b	6.75±0.25b
Moisture	(g/100g)	8.57±0.86ab	8.42±0.35a	8.57±0.02ab	8.45±0.07a
NFE	(g/100g)	36.28±0.26b	35.95±0.52a	38.00±0.04c	39.56±0.50d

D₁=Diet 1, D₂=Diet 2, D₃=Diet 3, D_c=Control diet, and NFE: Nitrogen free extract
a<b<c<d at (P<0.05)

similar ($P<0.05$) means in growth weight, daily growth rate and specific growth rate. However, fish that were fed on experimental diets exhibited a relative higher feed conversion ratio (FCR) compared to fish that were fed on control (commercial diet) (table 4). FCR did not differ significantly ($P<0.05$) among the diets that BSFLM substituted FM but was poor than the value recorded for fish fed on the control.

Experimental fish growth curves were exponential for the first five weeks of culture where fish growth assumed more or less linear trends for all the dietary treatments (Figure 2). Differential growth trends exhibited by experimental fish among dietary treatment occurred between weeks five and seven of culture. During the period of week five and seven, the growth curve of fish fed on control diet separated from the growth curves of fish fed on diets that BSFLM substituted FM (Figure 1). Growth appeared to slow down towards the end of the study period for fish subjected to control diet. The growth curves of the test diets had not separated much from each other is the reason of no statistical significance ($P<0.05$) observed for the diets. However, growth of fish fed on test diets appeared to rise exponentially towards the end of the culture period.

Water Quality Parameters

The experimental diets disturbed the water quality parameters contributing to marked significant difference ($P<0.05$) observed for dissolved oxygen (DO) and temperature between treatments (Table 5). However, pH, conductivity and total dissolved substance did not differ statistically ($P<0.05$) between the treatments. There was no significant difference ($P<0.05$) among water nutrients analyzed (table 5) except for phosphates that showed significant difference ($P<0.05$) between dietary treatments during the culture period (table 5).

Discussion

Presently, feeds nutritionists consider not only feed ingredients nutritional value but also price, availability on demand and sustainability for future use (Devic *et al.*, 2017). Constrains such as high cost, poor quality and unsustainable availability has forced feeds manufacturers to find alternative cost-efficient feeds ingredients to replace the more costly and commonly used FM (Gabriel *et al.*, 2007 and Liti *et al* 2005). Studies have shown insects proteins sources, e.g., BSFLM can be

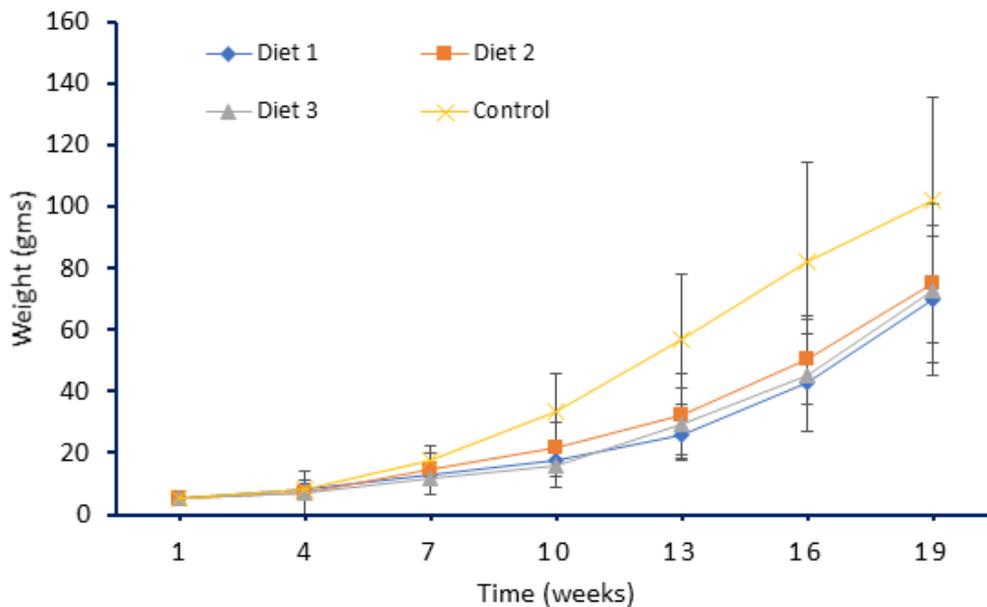


Figure 2. Growth curves for *C. gariepinus* under different BSFL dietary treatments (Diet 1; 50%) (Diet 2; 25%; (Diet 3; 75%) and (Diet 4; control) without BSFL

Table 4. Growth performance of *C. gariepinus* fed on diets containing varying percentages of BSFL and control diet.

BSFL (%)	Means ± SD						
	IW (g)	FW (g)	WG (g)	DGR	SGR	FCR	SR (%)
D1(50%)	5.1±0.00a	69.78±1.57a	64.09±1.57a	0.534±0.01a	2.17±0.01a	1.3±0.02a	100±0.00a
D2(25%)	5.1±0.00a	74.78±1.14b	69.78±1.14b	0.582±0.01a	2.24±0.15a	1.2±0.01a	100±0.00a
D3(75%)	5.1±0.00a	72.87±2.13b	67.77±2.13b	0.565±0.02a	2.22±0.02a	1.3±0.03a	98±0.57a
Dc (0%)	5.1±0.00a	102.17±1.41c	97.07±1.41c	0.809±0.02b	2.50±0.06a	1.1±0.01a	100±0.00a

IW: Initial weight, FW: Final weight, WG: Weight gain, DGR: SGR: Specific growth rate, Daily growth rate, FCR: Food conversion ratio, SR: Survival rate, D₁ =Diet 1, D₂ =Diet 2, D₃ =Diet 3 and D_c =Control diet; a<b<c<d at ($P<0.05$)

used as substitute to FM in aquaculture industry (Xiao *et al.*, 2018). However, dietary substitution of FM with BSFLM in aquafeeds remains controversial due to varying insects' nutritional quality, difference in diet formulation and fish species used in experiments (Matteo *et al.*, 2020). Substrates influence nutritional quality of BSFL biomass as a number of studies have demonstrated remarkable improvement when using substrates containing right polyunsaturated fatty acids (PUFAs) (Barroso *et al.*, 2017) as BSFL culture media. The crude protein percentage for BSFLM used for the present study was 25.3 % which is substantially lower compared to; (44.86-45.20) % used in trials for Rainbow trout (*Oncorhynchus mykiss*) by (Renna *et al.*, 2017), 41.74% reported by (Muin *et al.*, 2017) used in tilapia (*Oreochromis niloticus*) and 40% reported by (Newton *et al.*, 2005; Sheppard *et al.*, 1994) used in management of swine manure. According to (Barragan-Fonseca *et al.* 2017), crude protein content of BSFL decreases as age increases with highest percentage recorded for 5 days old larvae while the least percentage recorded for 20 days old larvae. As such, variation occurs in nutrients composition in the body of BSFL throughout the duration of larval development (Barragan-Fonseca *et al.* 2017) affecting growth rates of recipient organism e.g., fish when incorporated in diets depending on the stage of development. FCR is an important tool to determine efficiency, suitability and acceptability of formulated diets for fish (Nadaf *et al.*, 2010). Under optimal environmental condition and best pond management practices, high fish growth rate is achieved (USAID 2011) thus, better FCR. Other factors that influence FCR are type of fish; species and health status that contribute to variation in FCR. Under the current study, FCR of between 1.1 – 1.3 obtained among test diets were lower than FCR values obtained by (Tiamiyu *et al.*, 2017 and Amisah *et al.*, 2009) who tested *C. gariepinus* performance with *Allium sativum*, and *Leucaena leucocephala* replacement reported FCR values ranging between 5.71 – 6.32 and 4.95 – 6.39 respectively. The difference in FCR observed in various studies aforementioned may be attributed to feed ingredients variation, culture systems and varying stocking densities. The FCR under this study for *C. gariepinus* fed on 50% FM replacement indicate better growth

performance compared to 25% and 75% FM replacement diets. Similar results were reported by (Muin *et al.*, 2017) for Nile tilapia fingerlings fed on diet containing 50% FM replacement with FCR of 1.91. In another study by (St-Hilaire *et al.*, 2007), better growth performance with favorable FCR was reported on rainbow trout fed on 25% FM replacement diet. This can be explained by (Koprucu and Ozdemir 2005), who reported that BSFL contain chitin made of insoluble fiber that make digestion difficult for diets with high BSFL percentage hence affecting growth performance.

In this study, fish appeared healthy and there was no diseases outbreak throughout the trial period. All diets used for the trials were accepted by *C. gariepinus* irrespective of the percentage BSFL used in the substitution of FM paralleling reports by (Fawole *et al.*, 2019 and Adewolu *et al.*, 2010). The single mortality observed under the current study may have not been diet related but due to mishandling during sampling time thus, a higher survival rate (SR) was reported among the trial diets. The SR observed for this study was higher than that reported by (Nairuti *et al.* 2021) on *O. niloticus* but was similar to SR obtained by (Maina *et al.* 2020) on *C. gariepinus* under semi-intensive culture systems. Feed acceptance by experimental fish and good water quality parameters that were within the recommended range for fish culture (Tucker and Robinson 1990) may have contributed to the high SR mean. Uniform growth was observed irrespective of test diets subjected on experimental fish during the first three weeks of trials as shown in figure 1. The uniformity in growth may have been attributed to time taken for re-adjustment in digestion and nutrients uptake from experimental diets by *C. gariepinus* that were initially fed with wheat bran for a week before the trials. In support to our finding, (Kumar *et al.*, 2018) reported that the assimilation process of the digestive tract may be stimulated or impaired and therefore affect digestion on sudden change of diet and feeding regime in fish. It is during the initial stage of study that the slowest growth rates in *C. gariepinus* were observed in the entire trial duration. The trends observed during the initial stage of study were also obtained by (Maina *et al.* 2015) on *C. gariepinus* fingerlings subjected to diets containing 41% CP under the same FM replacement ratios as used

Table 5. Water quality parameters in tanks under different dietary treatments

Parameter	Unit	Means ± SD			
		D ₁	D ₂	D ₃	D _c
DO	(mgL ⁻¹)	3.03 ± 0.9 ^a	3.89 ± 0.74 ^c	3.66 ± 0.82 ^b	3.96 ± 0.46 ^d
Temperature	(°C)	24.53 ± 1.1 ^a	25.22 ± 1.27 ^b	25.98 ± 1.24 ^b	26.52 ± 1.65 ^c
pH		7.72 ± 0.52 ^b	7.72 ± 0.36 ^b	7.69 ± 0.54 ^a	7.72 ± 0.52 ^b
Conductivity	(µScm ⁻¹)	47.43 ± 3.39 ^b	45.9 ± 3.93 ^a	46.76 ± 3.45 ^b	46.81 ± 3.86 ^b
TDS	(mgL ⁻¹)	23.71 ± 1.89 ^b	22.86 ± 1.9 ^a	23.29 ± 1.68 ^b	32.3 ± 2.02 ^c
Ammonia	(mgL ⁻¹)	0.54 ± 0.51 ^a	0.61 ± 0.59 ^b	0.48 ± 0.46 ^a	0.71 ± 0.73 ^b
Nitrites	(mgL ⁻¹)	0.24 ± 0.15 ^a	0.22 ± 0.13 ^a	0.21 ± 0.12 ^a	0.30 ± 0.16 ^b
Nitrates	(mgL ⁻¹)	1.31 ± 0.78 ^b	1.0 ± 0.89 ^a	1.0 ± 0.92 ^a	1.16 ± 0.99 ^a
Phosphates	(mgL ⁻¹)	0.61 ± 0.07 ^c	0.52 ± 0.04 ^b	0.51 ± 0.1 ^b	0.47 ± 0.07 ^a

D₁, Diet 1; D₂, Diet 2; D₃, Diet 3; D_c Control Diet; DO, Dissolved oxygen; TDS; Total dissolved substance; a<b<c<d

in this study. Thereafter, growth curves of fish fed on control and diet containing (25%) BSFL separated from growth curves of fish fed on diets containing (50% and 75%) BSFL. The separation of the curves was a clear demonstration of better growth rates for control and 25% BSFL diets. However, control diet had the highest growth rate on fish as shown in figure 1 compared to the other test diets from the third week to the end of the trials. The trend observed for the control diet was also observed for 25% BSFL diet which was second in growth rate although not significantly different ($P>0.05$) compared to fish fed on (50% and 75%) BSFL test diets.

Fish fed on 50% BSFL had a better growth rate between week five and week eleven compared to fish fed on 75% BSFL which had the least growth rate for the first eleven weeks of the diets trial. After the eleventh week, the growth rate of 75% BSFL diet improved compared to 50% BSFL thus suggesting better growth performance as fish age on substitution of higher percentage of BSFL formulated diets. Final weight gain by fish fed on 50% BSFL diet was higher than fish fed on 75% BSFL diet paralleling results reported by (Steffens 2007 and Goda *et al.*, 2007) on rainbow trout and *C. gariepinus* having replaced FM by poultry by-products and insect meals respectively at 50% ratio. For present study, BSFL test diets were not supplemented with vitamins nor mineral premix that are essential for early-stage of fish development thus slow growth rate observed at the onset of the study. The slow growth rate observed at fingerling stage may be hasten by inclusion of necessary supplements which are either unavailable or are deficient in BSFL that was used in the feed trials. In intensive culture system where fish depend entirely on formulated feeds, the need for nutrients supplementation has been observed to increase with the variation between the standing fish biomass (De Silva 1992) and that of formulated food. Though the weight gains of fish fed on control diet was higher than for fish fed on BSFL diets, BSFL diets showed steady increase while control diet showed gradual slowing down in fish growth towards the end of the study. Therefore, BSFL formulated diets may outperform the control diet given more experimental time so long as the trends of curves towards the close of the trial are maintained. The weight gains of fish reported for feed trial under the present study shows that diets in which BSFL substituted FM obtained similar growth. The substitution percentages used in the present studies were similar to those used by (Fawole *et al.*, 2019 and Adeworu *et al.*, 2010) with results from both trial indicating BSFL can replace FM up to 75% without impairing and affecting performance in growth and survival of *C. gariepinus*.

Conclusion and Recommendation

This study established that dietary FM can be substituted by partially defatted BSFL ingredient of 25.3% CP at (25, 50 and 75) % rates without; variation in

growth performance and negatively affecting survival of *C. gariepinus* reared under aquaponic system. Thus, feed cost can substantially be reduced and profit margin increased by substituting dietary FM up to 75% for economic benefit in *C. gariepinus* production. Better growth rates for increased *C. gariepinus* productivity under aquaponic production system may be obtained by substitution of dietary FM using fully defatted BSFL with higher CP value above the 25.3 % used for the present study.

Ethical Statement

This study was carried out in accordance with the international, national and institutional guidelines for the care of experimental animals. No surgery was performed on the experimental organisms.

Funding Information

This experiment was funded by the World Bank through the government of Kenya (GoK) under the Kenya Climate Smart Agricultural Project (KCSAP).

Author Contribution

Conceptualization, B.M., R.K., K.O. and P.O.; Methodology B.M., R.K., K.O., P.O and J.M.; Investigation, B.M.; Formal analysis, B.M., R.K., K.O., P.O., J.M. and E.O.; Writing original draft, B.M.; Writing-reviewing and editing, B.M., R.K., K.O., P.O., J.M. and E.O.; Supervision, R.K., K.O. and P.O.; Project administration, KCSAP; Funding acquisition, B.M.

Conflict of Interest

The authors declare that they have no known competing financial or non-financial, professional or non-professional conflict that would have appeared to influence the work reported in this paper.

Acknowledgements

This research project was funded by the World bank through the Government of Kenya (GoK) under the Kenya Climate Smart Agricultural Program (KCSAP). The International Centre of Insect Physiology and Ecology (ICIPE) provided Black Soldier Fly Larvae that was used for the experiment. We acknowledge the department of Food Science and Technology of Jomo Kenyatta University of Agriculture and Technology (JKUAT) for providing Laboratory space and equipment for carrying out proximate analysis for the ingredients and test diets used for the experiment; Kenya Marine and Fisheries Research Institute (KMFRI) Baringo station for providing laboratory space and equipment for water quality analysis for nutrients. We thank Mr. David Chebon for having accepted the trials to be conducted on his farm in Mogotio, Baringo County, Kenya.

References

- Adeoluwa M.A., Ikenweibe N.B. and Mulero S.M. (2010) Evaluation of an animal protein mixture as a replacement for fish meal in practical diets for fingerlings of *Clarias gariepinus* (Burchell, 1822). *Israel Journal of Aquaculture-Bamidgeh* 62, 237-244
- Amisa S., Oteng M.A. and Ofori J.K. (2009) Growth performance of the African catfish (*Clarias gariepinus*) fed varying inclusion levels of *Leucaena leucocephala* leaf meal. *Journal of Applied Sciences and Environment Management* 13(1) 21-26 www.bioline.org.br/ja
- Al Mahmud N., Hasan M.D., Hossain M.B. and Minar M.H. (2012) Proximate composition of fish feed ingredients available in Lakshampur region, Bangladesh. *American-Eurasian Journal of Agriculture & Environmental Science* 12(5): pp. 556-560
- AOAC (Association of Official Analytical Chemistry) (1995) Official methods of analysis. 17th Edition. AOAC, Gaithersburg, MD, USA
- APHA (American Public Health Association) (1995) standard methods for the examination of water and wastewater. 19th Edition. APHA, Washington D.C., USA
- Badiola M., Basurko O.C., Piedrahita R.H., Hundley P. and Mendiola D., (2018) Energy use in recirculating aquaculture systems (RAS): A review. *Journal of Aquacultural Engineering* 81: pp. 57 – 70. <https://doi.org/10.1016/j.aqueng.2018.03.003>.
- Barbu M., Ceanga E. and Caraman S., (2016) Water quality modelling and control in recirculating aquaculture system; Intech Open Limited, The Shad 25th Floor, 32 London Bridge Street, London, SE1 9SG, United Kingdom. 53pp. DIO: 10.5772/63202
- Barragan-Fonseca K.B., Dicke M. and Van Loon J.J.A., (2017) Nutritional value of the black soldier fly (*Hermetia illucens* L.) and its suitability as animal feed- A review; *Journal of Insects as Food and Feed*, 3(2): 105-120
- Barroso F.G., Sanchez-Muros J.M., Segura M., Morote E., Ruiz A.T., Bueno R.R. and Guil-Guerrero J.L., (2017) Insects as food: Enrichment of larvae of *Hermetia illucens* with Omega 3 fatty acids by means of dietary modifications. *Journal of Food Composition and Analysis*. 62, 8-13. <https://doi.org/10.1016/j.jfca.2017.04.008>
- Chepkurui-Boit V., Ngugi C.C., Bowman J., Oyoo-Okoth E., Rasowo J., Bundi M.J. and Cherop L. (2011) Growth performance, survival, feed utilization and nutrients utilization of African catfish (*Clarias gariepinus*) larvae co-fed artemia and a micro-diet containing freshwater atyid shrimps (*Caridina nilotica*) during weaning, *Aquaculture Journal*, volume 17 issue 2 pages e82- e89. <https://doi.org/10.1111/j.1365-2095.2009.00737.x>
- De Silva S.S. (1992) Supplementary feeding in semi-intensive aquaculture systems. In farm made aquafeeds. Proceeding of FAO/AADCP, Bangkok, Thailand, pp. 24-60, FAO Rome Italy
- Devic E., Leschen W., Murray F. and Little D.C. (2017) Growth performance, feed utilization and body composition of advanced nursing Nile tilapia (*Oreochromis niloticus*) fed diets containing black soldier fly (*Hermetia illucens*) larvae meal. *Aquaculture Nutrition*. 1-8. <https://doi.org/10.1111/anu.12573>
- Ekawati W.A., Ulfa M.S., Yanuar T.A. and Kumiawan A. (2011) Analysis of aquaponic-recirculation aquaculture system (A- Ras) application in the catfish (*Clarias gariepinus*) aquaculture in Indonesia. *Aquaculture Studies*, 21, 93-100. http://doi.org/10.4194/2618-6381-v21_3_01
- Espinal C.A and Mutalic D., (2019) Recirculating aquaculture technologies: Aquaponic Food Production System; pp. 35-77. https://doi.org/10.1007/978-3-15943-6_3
- FAO (2014) Success factors in aquaculture enterprises in the Pacific: Farm Assets and Farm Performances of Private Aquaculture Enterprises, www.fao.org/contact-us/licence
- Fawole J.F., Adeoye A.A., Tihamiyu O.L., Ajala I.K., Obadara O.S. and Ganiyu O.I. (2019) Substituting fishmeal with hermetia illucens in diets of African catfish (*Clarias gariepinus*): Effects on growth, nutrient utilization, haemato-physiological response, and oxidative stress biomarker. *Aquaculture*. <https://doi.org/10.1016/j.aquaculture.2019.734849>
- FRDC (2001) A social-economic valuation of resource allocation options between recreational and commercial sectors: Economic Research Associates, Perth, Western Australia ISBN 0-9756020-0-4
- Gabriel U.U., Akinrotimi O.A., Bekibebe D.O., Onunkwo D.N., Anyanwu P.E. and Harcourt P. (2007) Locally produced fish feed: Potentials for aquaculture development in Sub-Saharan Africa. *Africa Journal of Agriculture*, 2, 287-295
- Goda A.M., El-Haroun E.R. and Kabir C. (2007) Effect of total or partial replacing fishmeal by alternative protein sources on growth of African catfish (*Clarias gariepinus*) (Burchell, 1822) reared in concrete tanks. *Aquaculture Resources*, 38(3): 279-287
- Huntington T., Avarre J.C., Focken U., Oswald M., Pucher J. and Sorgeloos P., (2017) Opportunities and challenges for aquaculture in developing countries; Joint Report by; Agence Francaise De Development (AFD), European Commission (EC) and Deutsche Gesellschaft for Internationale Zusammenarbeit (GIZ) <https://ec.europa.eu/development-cooperation-state-play-2015/en>
- Kim H. J., (2018) Aquaponics Basics: Horticulture and landscape architecture; Purdue University, Department of agriculture, pp. 6
- Kumar A., Pradhan P.K., Chadha N.K., Mohindra V., Tiwari V.K. and Sood N. (2018) Effects of dietary regimes on development of digestive system of stinging catfish (*Heteropneustes fossilis*, Bloch) larvae. *International Journal of Current Microbiology and Applied Science*. ISSN: 2319-7706: 7(8): 413-421
- Kopruca K. and Ozdemir Y. (2005) Apparent digestibility of selected feed ingredients for Nile tilapia (*Oreochromis niloticus*). *Aquaculture*, (250), 308-316
- Liti D., Cherop L., Munguti J. and Chhorn L., (2005) Growth and economic performance of Nile tilapia (*Oreochromis niloticus* L.) fed on two formulated diets and two locally available feeds in fertilized ponds: *Aquaculture Research*, Vol 36 pp. 746-752. <https://doi.org/10.1111/j.1365-2019.2005.01265.x>
- Liti D. M., Mugo R. and Muchiri M. (2002) Growth and economic performance of Nile tilapia fed on three brans in fertilized ponds: *Aquaculture America*, California USA, 2002, pp. 187
- Love C.D., Fry P.J., Li X., Hill S E., Genello L., Semmens K., Thompsons K. (2015) Commercial aquaponics production and profitability. Finding from an international survey: *Aquaculture Journal*, (435) 67-74
- Maina A.N., Osugo I.M., Munga L.K. and Munguti J.M. (2020) Growth performance and carcass characteristics of the

- African catfish (*Clarias gariepinus*) reared on diets containing black soldier fly (*Hermetia illucens*) larval meal. MSc Thesis, Kenyatta University
- Matteo Z. Basilio R., Giorgia G., Christina T., Elisabetta G., Paola R., Giorgia G., Cristiona B., Andrea O., Gloriana C., Tyrone L., Vesna M., Anna A., Francesca T., Valentina N., Sara R., Francesca C. and Like O. (2020) Zebra (Danio rerio) physiological and behavioural responses to insect-based diets. A multidisciplinary approach. Scientific reports, Nature Research.
<https://doi.org/10.1038/s41598-020-67740-w>.
- Miles D.R. and Jacob P.J. (2011) Fishmeal: Understanding why this feed ingredient is so valuable in poultry diets; Animal science department, Florida cooperative extension services, institute of food and agricultural science, university of Florida, (1-3) <http://edis.ifas.ufl.edu>.
- Muin H., Taufek N.M., Kamarudin M.S. and Razak S.A (2017) Growth performance, feed utilization and body composition of Nile tilapia, *Oreochromis niloticus* (Linnaeus, 1758) fed with different levels of black soldier fly, *Hermetia illucens* (Linnaeus 1758) maggot meal diet. Iranian Journal of Fisheries Science 16(2) 567-577
- Munguti J., Obiero K., Odame H., Kirimi G. J., Kyule D., Josiah A. and Liti D. (2021) Key limitation of fish feeds, feed management practices, and opportunities in Kenya's aquaculture enterprise, African Journal of Food, Agriculture, Nutrition and Development 21(21):17415-17434.
<https://doi.org/10.18697/ajfand.97.20455>
- Munguti, J.M. Safina M., Orina P.S., Kyule D.N., Opiyo M.A., Karisa H.C. and Ogello E.O. (2014) An overview of current status of Kenyan fish feed industry and feed management practices, challenges and opportunities: International Journal of Fisheries and Aquatic studies. Vol 1 no 6 pp. 128-137
- Munguti J.M., Waidbacher H., Liti D.M., Straif M. and Zollitsch W. (2009) Effect of substitution of freshwater shrimp meal (*Caridina nilotica* Roux) with hydrolysed feather meal on growth performance and apparent digestibility in Nile tilapia (*Oreochromis niloticus* L.) under different culture conditions. Livestock Research for Rural Development, 21, 1-13
- Nadaf S.B, Bhilave M.P. and Deshapande V.Y. (2010) Growth performance and feed conversion ratio of freshwater fish fed on formulated feed. Journal of Aquaculture Biology, 25(1): 181-185
- Nairuti R.N., Munguti J.M., Waidbacher H. and Zollitsch W. (2021) Growth performance and survival rate of Nile tilapia (*Oreochromis niloticus* L.) reared on diets containing black soldier fly (*Hermetia illucens* L.) larval meal. Journal of Land Management, Food and Environment. Volume 72 Issue 1, 9-19
<https://doi.org/10.2478/boku-2021-0002>
- Naylor L.R., Goldburg J.R., Primavera H.J., Kautsky N., Beveridge M.C.M., Clay J., Folke C., Lubchenco., Mooney H. and Troell M. (2000) Effects of aquaculture on world fish supplies; Nature, volume 405 1017-1024
<https://doi.org/10.1038/35016500>
- Newton L., Sheppard C., Watson D., Burtle G. and Dove R. (2005) Using the black soldier fly *Hermetia illucens*, as a value-added tool for the management of swine manure. Animal and poultry waste management center, North Carolina State University, Raleigh, NC 17
- Ngugi C. C., Bowman J. R. and Omolo B. O., (2007) A new guide to fish farming in Kenya. Aquaculture collaborative research support program (ACRSP) management Office, Oregon State University, Oregon USA. 100pp.
http://pdacrsp.oregonstate.edu/pubs/featured_titles/kenya_Manual.pdf
- Omondi, J.G; Gichuri, W.M and Veverica, K. (2001) A partial economic analysis for Nile tilapia (*Oreochromis niloticus*) and sharp toothed African catfish (*Clarias gariepinus*) polyculture in central Kenya. Blackwell journal of Aquaculture Research Vol 32: pp. 693-700
- Renna M., Schiavone A., Gai F., Dabbou S., Lussiana S., Malfatto V., Prearo M., Capucchio M.T., Biassato I., Biasibetti E., De Marco M., Brugiapaglia A., Zoccarato I. and Gasco L. (2017) Evaluation of the suitability of partially defatted (*Hermetia illucens* L.) Larvae meal as ingredient for rainbow trout (*Oncorhynchus mykiss* Walbaum) diets. Journal of Animal Science and Biotechnology.
<https://doi.org/10.1186/s40104-017-0191-3>
- Rurangwa E. and Verdegem M.C.J., (2014) Micro-organisms in recirculating aquaculture systems and their management, Reviews in Aquaculture, INASP KENYA-Karatina university College;
<https://doi.org/10.1111/raq.12057>
- Safran P., (2009) Fisheries and Aquaculture; Fishing in the tropics, Philippine council for aquatic and marine research and development, Los Banos Laguna, Philippines Vol IV pp. 134-154
- Sheppard D.C., Newton G.L., Thompson S.A. and Savage S. (1994) A value-added manure management system using black soldier fly. Bioresource Technology 502 275-279
- Somerville C., Cohen M., Pantanella E., Stankus A. and Lovatelli A., (2014) Small-scale aquaponic food production system; integrated fish and plant farming; FAO Fisheries and Aquaculture Technical Paper 589
- Stamer A., Wessels S., Neidigk R. and Hoersten G. (2014) Black soldier fly (*Hermetia illucens*) larval-meal as an example of a new feed ingredients' class in aquaculture diets. Rahmann G and Aksoy U (Eds). (2014) Proceedings of the 4th ISOFAR Scientist conference. 13-15 Oct, Istanbul, Turkey (e-print ID 24223)
- Steffens W. (1994) Replacing fishmeal with poultry by-product meal in diets for rainbow trout (*Oncorhynchus mykiss*). Aquaculture, 124: 27-34
- St-Hilaire S., Sheppard C., Tomberlin J.K., Irving S., Newton L., McGuire M.A., Mosley E.E., Hard R.W. and Sealey W. (2007) Fly prepupae as a feedstuff for rainbow trout (*Oncorhynchus mykiss*). Journal of the World Aquaculture Society, 38: 59-67
- Tiamiyu A.M., Adedeji O.B. and Olatoye I.O. (2017) Growth performance of the African catfish (*Clarias gariepinus*) fed varying inclusion of *Allium sativa* as feed additives. American Journal of Biotechnology and Bioinformatics 1(1): 1-7 ISSN:257599X
- Tilami S.K., Turek J., Cervery D., Lepic P., Kozak P., Burkina V., Sakallis S., Tocala A., Sampels S. and Mraz J. (2020) Insect meal as a partial replacement for fishmeal in a formulated diet for perch (*Perca fluviatilis*), Turkish Journal of Fisheries and Aquatic Sciences. 20(12): 867-878
- Tran G., Gnaedinger C. and Merlin C., (2015) Black soldier fly larvae (*Hermetia illucens*). Feedipedia, a programme by INRA, CIRAD, AFZ and FAO.
<https://www.feedipedia.org/node/16388>
- Tucker C.S and Robinson E.H. (1990) Water Quality

- Management in Ponds. In: channel catfish farming handbook. Springer, Boston, MA.
https://doi.org/10.1007/978-1-4757-1376-3_10
- USAID (2011) Harvest. Helping address rural vulnerability and ecosystem stability. Technical Bulletin, 7: 1-2
www.cambodiaHARVEST.org
- Walker A.B., Fournier H.R., Neefus C.D., Nard G.C. and Berlinsky D.L. (2009) Partial replacement of fishmeal with Larval Pophyra Spp. In diets for Atlantic cod. North American Journal of Aquaculture. 71, 39-45
- Xiao X., Jin P., Cai M., Yu Z., Yu J. and Zheng J., (2018) Effects of black soldier fly (*Hermetia illucens*) larvae meal protein as a fishmeal replacement on the growth and immune index of yellow catfish (*Pelteobagrus fulvidraco*); Aquaculture Research. 2018 pp. 1-9
<https://doi.org/10.1111/are.13611>